

## Technical Data Sheet

# CE Bismuth Sulfite Agar acc. to Wilson-Blair

Ordering number: 1.05418.0500

Bismuth Sulfite Agar acc to Wilson-Blair is a selective agar introduced by Wilson and Blair (1927, 1931) for the isolation and differentiation of *Salmonella typhi* and other *salmonellae* from clinical specimens, e.g. feces.

IVD in vitro diagnosticum - For professional use only

### Mode of Action

Brilliant green and bismuth largely inhibit the accompanying bacterial flora. Colonies of H<sub>2</sub>S-positive *salmonellae* exhibit blackening due to the formation of iron sulfide. Reduction of bismuth ions to metallic bismuth produces a metallic lustre around the colonies (McCoy 1962).

Bismuth sulfite agar is highly selective and a preferred medium for the isolation of *Salmonella typhi*. *Salmonella typhi* is more frequently isolated from blood cultures than from fecal specimens. Blood cultures are positive for 80% of typhoid patients during the first week of fever but show decreasing positive results thereafter. Gram-positive bacteria and coliforms are inhibited on Bismuth Sulfite Agar. It is a standard methods medium for the clinical environment, industrial applications and it is accepted for routine detection of most *Salmonella* spp. The freshly prepared medium is strongly inhibitory and is suitable for heavily contaminated samples.

### Typical Composition

Meat Extract	5 g/l
Peptone from Meat	5 g/l
Peptone from Casein	5 g/l
D(+)-Glucose	5 g/l
K <sub>2</sub> HPO <sub>4</sub>	4 g/l
Iron(III) Sulfate	0.3 g/l
Brilliant Green	0.025 g/l
Bismuth Sulfite Indicator	8.5 g/l
Agar-Agar	15 g/l

## Preparation

Suspend 47.5 g/l. Mix the resulting precipitate to give a uniform suspension. Pour plates to give thick layers (25 ml). **Do not autoclave.**

The appearance of the prepared medium is turbid and green.

The pH at 25 °C is in the range of 7.4 -7.8.

The freshly prepared medium is strongly inhibitory and is thus especially suitable for heavily contaminated specimen, e.g. feces.

Acc. to FDA-BAM the medium should be prepared 1 day before use, store dark. Loss of selectivity after 48 h.

## Specimen

e.g. Stool.

Clinical specimen collection, handling and processing. See general instructions of use.

## Experimental Procedure and Evaluation

Inoculate by thinly spreading the sample or material from an enriched culture on the surface of the medium.

Incubation: up to about 48 h at 35 °C aerobically.

*Salmonella* colonies often display blackening after 18 h of incubation, the metallic sheen appears several hours later depending on the age of the medium.

Appearance of Colonies	Microorganisms
Black colonies with sheen surrounded by brownish-black zones	<i>Salmonella typhi</i>
Black or greenish-gray colonies, may have sheen, with or without darkening of the surrounding medium	<i>Salmonella</i> spp.
Small, green to brown colonies, sometimes mucoid	<i>Coliform bacteria</i> , <i>Proteus</i> and others

## Limitations

1. It is important to streak for well isolated colonies.
2. With confluent growth typical colonial characteristics of *Salmonella* spp. will not develop
3. Some *Salmonella* strains are markedly inhibited, for example *Salmonella gallinarum*, *Salmonella sendai*, *Salmonella berta*, *Salmonella abortus-equi* (Hajna 1951)

Therefore, when in doubt, almost any growth on the medium should be subject to further tests e.g. subculture onto a less selective medium in a manner to obtain well-isolated colonies. Use pure cultures for biochemical and serological confirmation.

## Storage

The product can be used for sampling until the expiry date if stored upright, protected from light and properly sealed at +15 °C to +25 °C.



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After first opening of the bottle the content can be used up to the expiry date when stored dry and tightly closed at +15 to +25° C.

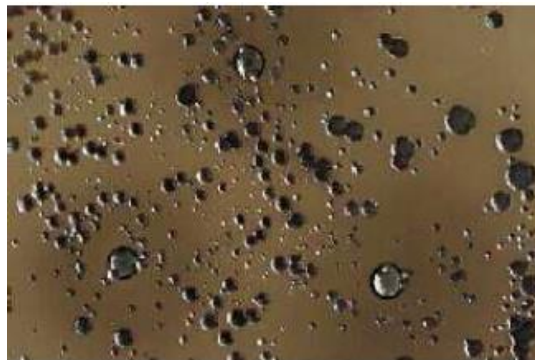
## Disposal

Please mind the respective regulations for the disposal of used culture medium (e.g. autoclave for 20 min at 121 °C, disinfect, incinerate etc.).

## Quality Control

Control Strains	ATCC #	Incubation	Expected Results
<i>Salmonella typhimurium</i>	14028	48 h at 35 °C	Growth good to very good, black colonies, metallic lustre
<i>Salmonella typhimurium</i>	13311	48 h at 35 °C	Growth good to very good, black colonies, metallic lustre
<i>Salmonella enteritidis</i>	5188	48 h at 35 °C	Growth good to very good, black colonies, metallic lustre
<i>Salmonella arizonae</i>	13314	48 h at 35 °C	Growth fair to very good, black colonies, metallic lustre
<i>Salmonella aboni</i>	6017	48 h at 35 °C	Growth good to very good, black colonies, metallic lustre
<i>Escherichia coli</i>	25922	48 h at 35 °C	Growth poor to fair, green colonies, no metallic lustre
<i>Proteus mirabilis</i>	19906	48 h at 35 °C	Growth good to very good, olive colonies, no metallic clustre
<i>Shigella sonnei</i>	11060	48 h at 35 °C	No growth
<i>Staphylococcus aureus</i>	25923	48 h at 35 °C	No growth
<i>Bacillus cereus</i>	11778	48 h at 35 °C	No growth

Please refer to the actual batch related Certificate of Analysis.



*Salmonella typhimurium*

## Literature

American Public Health Association (1992). Compendium of Methods for the Microbiological Examination of Foods. 3<sup>rd</sup> edition.

Hajna, A. A. (1951). Preparation and application of Wilson and Blair's bismuth sulfite agar medium. The Public Health Laboratory, 9: 48-50.

McCoy, J.H. (1962). The isolation of *Salmonellae*. J. Appl. Bact. **25**: 213-224.



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Wilson, W.J. and Blair, E.M. (1927). McV.: Use of glucose bismuth sulfite iron medium for the isolation of *Bacillus typhosus* and *Bacillus proteus*. J. Hyg., **26**: 374-391.

Wilson, W.J. and Blair, E.M. (1931). McV.: Further experience of the bismuth sulfite media in the isolation of *Bacillus typhosus* and *Bacillus paratyphosus* B from faeces, sewage and water. J. Hyg. **31**: 138-161.

### Ordering Information

Product	Cat. No.	Pack size
Bismuth Sulfite Agar acc. to Wilson-Blair	1.05418.0500	500 g



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