

Distribué par :

Z.A de Gesvrine - 4 rue Képler - B.P.4125 44241 La Chapelle-sur-Erdre Cedex - France t.: +33 (0)2 40 93 53 53 | f.: +33 (0)2 40 93 41 00 commercial@humeau.com



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# MC-Media Pad

# A Convenient and Approved Method for Counting Contaminants in Food and Beverage Products

The MC-Media Pad is a convenient method for rapid routine testing of raw materials and in-process testing of finished food and beverage products for microbial contamination.

The MC-Media Pad solution is composed of a series of ready-to-use pads for total count and specific detection and enumeration of indicator organisms. MC-Media Pads target 4 key applications in food and beverage microbial testing (Table 1). The MC-Media Pad method has been independently tested and evaluated. Media Pad Rapid Aerobic Count (RAC), MC-Media Pad Coliform, MC-Media Pad E. coli & Coliform and MC-Media Pad Yeast & Mold received certification from AOAC Research Institute as a Performance Tested MethodSM<sup>(1, 2, 3, 4)</sup>.

Table 1: MC-Media Pad product solution for food and beverage applications

Product Name	Application
MC-Media Pad Rapid Aerobic Count	Enumeration of aerobic microbial contamination
MC-Media Pad Coliform	Enumeration of coliform bacteria
MC-Media Pad <i>E. coli</i> & Coliform	Simultaneous enumeration of Escherichia coli and coliform bacteria
MC-Media Pad Yeast & Mold	Enumeration of total yeast and mold count

This document will focus on results obtained for the 4 certified MC-Media Pads: MC-Media Pad RAC, MC-Media Pad Coliform, MC-Media Pad *E. coli* & Coliform and MC-Media Pad Yeast & Mold.

# **Technology and principle**

MC-Media Pad technology is based on a fabric pad placed on an adhesive support and covered with a transparent gas permeable lid. The fabric pad is a multi-layered structure composed of a water-soluble high polymer laminated with porous non-woven fabric. Dedicated formulations of culture media incorporating nutrients and selective agents are coated onto the fabric pad. The MC-Media Pad RAC promotes the growth of aerobic bacteria in red colonies. The MC-Media Pad Coliform nutrient film also contains X-Gal (5-bromo-4-chloro-3-indoxyl-βD-galactopyranoside) which is hydrolyzed by  $\beta$ -galactosidase in coliforms to produce blue colonies. The MC-Media Pad E. coli & Coliform, in addition to X-Gal, contains Salmoglucuronic acid (6-chloro-3-indoxyl-βD-glucuronic acid) which is hydrolyzed by  $\beta$ -glucuronidase in *E. coli* to specifically stained E. coli colonies purple. MC-Media Pad Yeast & Mold promotes the growth of yeasts and molds in red colonies.

The fabric pad has been specifically developed for the testing of 1 mL food samples. Sample solution diffuses throughout the entire pad to dissolve and release the nutrient compound forming a highly viscous solution.

The combination of the coated pad and the gas permeable lid promotes the selective or universal growth of microorganisms and enables enumeration.

MC-Media Pad RAC, MC-Media Pad Coliform, MC-Media Pad *E. coli* & Coliform and MC-Media Pad Yeast & Mold performances have been challenged with a range of relevant food matrices either naturally or artificially contaminated with indicator organisms.

# **Method**

# **Sample Preparation**

Portions of 50 g, 25 g or 25 mL of each food sample were added to a stomacher bag or blender cup. 450 mL or 225 mL of the relevant diluent (Peptone water, Butterfield's phosphate buffer, sterile physiological saline or sodium citrate) was then added to the stomacher bag or blender cup and stomached or homogenized for 2 min. Decimal dilutions can be performed depending on the sample.

# **MC-Media Pad protocol**



Remove MC-Media Pad from aluminium pouch



Label MC-Media Pad Open the cover film diagonally



Dispense 1 mL of food sample onto the center of the pad. Sample diffuses automatically to whole pad



Close cover film and incubate according to application conditions



After incubation, count colonies in CFU

# **Results**

# **MC-Media Pad RAC**

Total aerobic enumeration has been determined for the 10 following food matrices: Cooked, peeled, chilled cold water prawns (1.3% salt); prewashed bagged flat leaf parsley; chilled tuna pate (23% fat, 0.9% salt); fermented strawberry yogurt drink (1.8% fat containing *Lactobacillus acidophilus* and *Bifidobacterium* and *Lactobacillus casei*); fresh raw chicken breast fillets (2% fat); fresh raw lean pork mince (5% fat); cold press green vegetable juice (cucumber, celery, spinach, lemon, lettuce, pear, pineapple); full fat soft cream cheese (24% fat), fresh chilled egg mayonnaise sandwish on white sliced bread (5% fat, 0.9% salt) and fresh feta cheese deli pasta salad (3.4% fat, 0.5% salt).

Rapid detection incubation (35 °C, 24 h) and standard detection incubation (35 °C, 48 h) conditions were used. The standard incubation condition is applicable for all food matrices but is more appropriate when food samples contain psychrophilic bacteria or a large amount of acidic bacteria since they tend to grow slowly at non-optimum temperatures. Among the 10 food matrices, 7 were selected to compare the performance in the rapid detection incubation condition versus the reference method.

Total aerobic bacteria was detected after 24 hours of incubation at 35 °C for the 7 matrices and after 48h at 35 °C for the 10 matrices.

In a separate study, all matrices were compared to the ISO 4833:2013 method with an incubation of 72 h at 30  $^{\circ}$ C. Total aerobic bacteria was detected after 72 h of incubation at 30  $^{\circ}$ C for the 10 matrices



All grown colonies develop reddish color

# Regulatory approval

MC-Media Pad RAC was certified by AOAC Research Institute as a Performance Tested Methods<sup>SM</sup> Program.

Certificate number is 091702.

# **MC-Media Pad Coliform**

# Inclusivity and exclusivity tests

Inclusivity and exclusivity tests have been performed on 48 coliform and 39 non-coliform type strains including some food isolates.¹ BGLB broth was used as control. MC-Media Pad Coliform and BGLB broth were incubated 24 h at 35 °C.

All coliform strains on MC-Media Pad showed positive results with a blue-green staining of each colony. The results indicate an inclusivity rate of 100%.

Results for all non-coliform strains were negative both in the BGLB and MC-Media Pad Coliform indicating an exclusivity rate of 100%.<sup>1</sup>

#### Food matrices tested

Detection of coliforms in 26 food samples have been performed. Food types belong to 9 food categories: meat, poultry, fish and seafood, fruits and vegetable, dairy, chocolate and bakery, animal feeds, pasta.

Results showed a detection of coliforms after 24 hours of incubation at 35  $^{\rm o}{\rm C.^1}$ 



Coliforms develop blue/blue-green colored colonies due to  $\beta$ -galactosidase production. Gram negative non-coliform bacteria form colorless colonies.

#### Regulatory approval

MC-Media Pad Coliform was certified by AOAC Research Institute as a Performance Tested Methods<sup>SM</sup> Program.

The certificate number is 100402.

# MC-Media Pad E. coli & Coliform

# Inclusivity and exclusivity

Inclusivity and exclusivity tests have been performed on 24 strain types of *E. coli*, 8 *E. coli* strains isolated from foods, 21 coliform strain types other than *E. coli*, and 12 coliform strains from food 2, and 65 non-coliform strains were added in the study. BGLB broth was used as the control. MC-Media Pad *E. coli* & Coliform and BGLB broth were incubated 24 h at 35 °C.

Strains showing gas production in BGLB grew as coliform blue colonies on MC-Media Pad  $E.\ coli$  & Coliform. All  $E.\ coli$  strains produced a purple to indigo staining on MC-Media Pad. The results indicate an inclusivity rate of  $100\%.^2$ 

Results of all non-coliform strains were negative both in the BGLB and MC-Media Pad *E. coli* & Coliform except *Aeromonas hydrophila, Serratia marcescens* and *Serratia rubidae* which showed a positive result on MC-Media Pad indicating an exclusivity rate of 95.4%.<sup>2</sup>

#### **Food Matrices tested**

Detection of coliforms and *E. coli* has been performed on 100 samples of meats and meat products.

Results showed a detection of coliforms and *E. coli* after 24 hours of incubation at 35 °C.<sup>2</sup>



Coliforms develop blue/blue-green colored colonies due to  $\beta$ -galactosidase production whereas  $E.\ coli$  form indigo to purple colored colonies due to specific  $\beta$ -glucuronidase for  $E.\ coli$ . Gram-negative non-coliform bacteria form colorless colonies.

# Regulatory approval

MC-Media Pad *E. coli* & Coliform was certified by AOAC Research Institute as a Performance Tested Methods<sup>SM</sup> Program.

The certificate number is 070901.

# MC-Media Pad Yeast & Mold

# Comparison of MC-Media Pad Yeast and Mold and FDA/BAM reference method

Yeast and mold detection has been performed on contaminated food and artificially contaminated samples, specifically orange juice concentrate, cake mix, breaded chicken nuggets, dry pet food and yogurt.

Products were inoculated at 3 inoculation levels: low (102 CFU/g), medium (103 CFU/g) and high (104 CFU/g).

DG18 agar was used a control for foods with high water activity ( $a_w$ >0.95), and DRBC agar was used as control for low water activity food samples ( $a_w$ <0.95).

Results showed a detection of yeasts and molds after 48 to 72 hours of incubation at 25 °C.<sup>3</sup>



All grown colonies develop a reddish color. Yeasts appear as circular reddish colored colonies, whereas molds appear as diffuse and fuzzy round, reddish colored colonies.

#### Regulatory approval

MC-Media Pad Yeast & Mold was certified by AOAC Research Institute as a Performance Tested Methods<sup>SM</sup> Program.

The certificate number is 111401.

# **Conclusion**

This application study shows that the AOAC certified MC-Media Pad RAC, MC-Media Pad Coliform, MC-Media Pad *E. coli* and Coliform and MC-Media Pad Yeast and Mold can be used to detect and easily enumerate indicator organism contamination in a broad range of food and beverage matrices.

Chromogenic indicators and dedicated culture media produce specific results and allow ease of interpretation. Additionally providing the benefits of a convenient media method through a well-defined inoculation area and reduction of required space for storage, incubation and waste.

#### References:

- Morita H, Ushiyama M, Aoyama S and Iwasaki M. Journal of AOAC International Vol. 89, N° 2, 2006.
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- Caballero O, Alles S, Le Q, Mozola M and Rice J. Journal of AOAC International Vol. 98, N° 3, 2015.
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